$See \ discussions, stats, and author \ profiles \ for \ this \ publication \ at: \ https://www.researchgate.net/publication/312317617$

Novel insight on probiotic Bacillus subtilis: Mechanism of action and clinical applications

Article · January 2017

CITATIONS		READS
3		1,807
1 autho	:	
R	Manoj Suva	
	37 PUBLICATIONS 77 CITATIONS	
	SEE PROFILE	
_	the suthers of this publication are also working on these related projects	

Some of the authors of this publication are also working on these related projects:

Aflapin (Boswellia serrata) for arthritis management View project



Pharmacovigilance View project

Review Article

Novel insight on probiotic *Bacillus* subtilis: Mechanism of action and clinical applications

Manoj A. Suva, Varun P. Sureja, Dharmesh B. Kheni

Department of Pharmacology, K. B. Institute of Pharmaceutical Education and Research, Kadi Sarva Vishwavidyalaya University, Gandhinagar, Gujarat, India

Abstract Probiotics are the living microorganisms that provide health benefits to the recipient. *Lactobacillus* and *Bifidobacterium* genera have been used since long for the competitive exclusion of pathogens from the gut. However, their limitations such as sensitivity to gastric acid, temperature, slow growth, and specific stability conditions lead to search for a novel probiotic that is stable through its shelf-life as well as during gastrointestinal transit; hence, offering better efficacy. *Bacillus* bacteria have strong scientific data which substantiates the validity of the use as preferred probiotics. In recent times, there has been significant progress in scientific evaluation and studies on probiotic *Bacillus subtilis*, revealing possible mechanisms of action like antimicrobial effect by synthesis of antimicrobial substances, antidiarrheal effect, immunostimulatory effect, competitive exclusion of pathogens, prevention of intestinal inflammation, and normalization of intestinal flora. Numerous preclinical and clinical studies on *B. subtilis* have shown its promising efficacy in the treatment and prevention of diarrhea of various etiologies. *B. subtilis* is certified as generally recognized as safe by Food and Drug Administration and features in the European Food Safety Authority Qualified Presumption of Safety, hence suggesting as safe for human use. All of these beneficial attributes make *B. subtilis* the most attractive probiotic species for various clinical conditions.

Key Words: Antidiarrheal effect, antimicrobial substances, Bacillus subtilis, immunomodulation

Address for correspondence:

Mr. Manoj A. Suva, Department of Pharmacology, K. B. Institute of Pharmaceutical Education and Research, Kadi Sarva Vishwavidyalaya University, Gandhinagar - 382 024, Gujarat, India. E-mail: manojsuva_0211@yahoo.co.in Received: 22.10.2016, Accepted: 18.11.2016

INTRODUCTION

The term "probiotics" was derived from the Greek word, meaning "for life."^[1] According to the WHO/FAO, probiotics are: "Live microorganisms which when administered in adequate amounts confer a health benefit on the host."^[2] Various factors such as environmental, nutritional and/or metabolic changes favor pathogen proliferation and that disturb equilibrium between beneficial and pathogenic flora leading to disease

Access this article online				
Quick Response Code:	Website:			
	http://www.jcrsmed.org			
	DOI: 10.4103/2455-3069.198381			

conditions. Synthetic antibiotics were the first option to control pathogenic overgrowth; however, the unregulated use of these compounds induced multi-drug resistance in pathogenic bacteria. Hence, antibiotic utilization needs to be well regulated. In this sense, the utilization of beneficial bacteria (probiotics) has emerged as an alternative based on the beneficial good results obtained with it.^[3] Lactobacillus

For reprints contact: reprints@medknow.com

How to cite this article: Suva MA, Sureja VP, Kheni DB. Novel insight on probiotic *Bacillus subtilis*: Mechanism of action and clinical applications. J Curr Res Sci Med 2016;2:65-72.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

and *Bifidobacterium* genera have been used almost exclusively for the competitive exclusion of pathogens from the gut of humans and animals. However, there are some major challenges related to manufacturing probiotic formulations containing lactobacilli and *Bifidobacteria spp.*, as below:

- These microorganisms are microaerophilic or strict anaerobic, therefore, their handling and production is complex and could be a challenge
- Both microorganisms are slow growers
- Sensitive to temperature hence product must be maintained at low temperatures and shelf-life in general is short
- Many of lactobacilli and *Bifidobacteria* are sensitive to gastric juice during gastrointestinal (GI) tract transit.^[4]

Bacillus bacteria are highly diverse group of microorganisms, known for more than 100 years. There are strong scientific data which substantiates the validity of the use of *Bacillus* bacteria as probiotics.^[5] Only a few of the 100 species contained in the *Bacillus* genus are used as probiotics in humans such as *Bacillus subtilis, Bacillus licheniformis, Bacillus coagulans, Bacillus cereus var. toyoi, Bacillus natto (subtilis), Bacillus clausii, Bacillus pumilus, and <i>Bacillus cereus*.^[6] In recent decades, scientific studies on probiotic *B. subtilis* have made significant progress to elucidate the activity spectrum which makes this bacterium the most attractive probiotic for clinical use. In this review, we have present data from experimental and clinical research that may allow making an impression of the therapeutic potential of *B. subtilis*.

BACILLUS SUBTILIS BACTERIA IN NATURE

Bacillus constitutes a diverse group of rod-shaped, Gram-positive bacteria, characterized by their robust spore-forming ability.^[3] *Bacillus* genus bacteria are most widespread microorganisms in nature. *Bacillus* species are predominant in soil and also have been isolated from water, air, and food products like wheat, grain, wholemeal, soya beans, and milk microflora. Bacilli consistently enter the GI and respiratory tracts of healthy people with food, water, and air. Isolation of *B. subtilis* from the human GI tract was reported for healthy adults and children. The number of bacilli in the gut can reach 10⁷ CFU/g, comparable to lactobacilli count. Thus, researchers considered *Bacillus* to be one of the dominant components of the normal gut microflora.^[5] The *B. subtilis* genome is totally sequenced, leading to the generation of great amount of basic knowledge and also developments of molecular and genetic methodologies.^[3]

BACTERIAL SPORE FORMATION

Bacterial spores are formed as a means to survive extreme environmental conditions for long-term survival otherwise that conditions kill vegetative bacteria. Sporulation is very



Figure 1: Sporulation life cycle of bacterial spore formers

much dependent upon the nutrients availability in the immediate vicinity of the live cell. According to Cutting *et al.* (2009), decrease in nutrients is sensed by the bacterium and it enters an irreversible program of development that results in the production of spores as shown in Figure 1.^[7] Bacterial endospore contains a condensed and inactive chromosome at its core. Surrounding the spore, layer of peptidoglycan-rich cortex and one or more layers of proteinaceous material is present referred as the spore coat. During the lack of nutrient condition, growing vegetative cell (VC) undergoes a series of morphological changes that forms a forespore (F) within the mother cell (MC) of the sporangium and then after some time spore (S) is released by lysis of the MC.^[7]

BACILLUS SUBTILIS SPORE FORMATION AND ITS ADVANTAGES

B. subtilis spores are highly resistant to temperature, extreme pH, gastric acid; bile and solvents, hence keep viability in the gut.^[3,5] *B. subtilis* can be stored for long time periods without refrigeration.^[3]

BACILLUS SUBTILIS SPORE GERMINATION AND PROLIFERATION IN GASTROINTESTINAL TRACT

Immunological data showed that *B. subtilis* spores germinate in the mouse gut and proliferate and able to grow and resporulate.^[8] Casula and Cutting observed germination of *B. subtilis* spores as VCs in the mouse jejunum and ileum using a competitive reverse transcription polymerase chain reaction targeting a genetically engineered chimeric gene, ftsH-lacZ, which is only expressed by VCs.^[9] Ozawa *et al.* found that spores of *B. subtilis* strain BN germinate and multiply to some extent in the GI tract of pigs.^[10] Leser *et al.* showed that *B. subtilis* germinate and grow in proximal part of the pig GI tract.^[11]

MECHANISM OF ACTION OF PROBIOTIC BACILLUS SUBTILIS

Possible mechanisms of action include antimicrobial effect by synthesis of antimicrobial substances, antidiarrheal effect, immunostimulatory effect, competitive exclusion of pathogens, prevention of intestinal inflammation, and stimulation of growth of intestinal normal flora.^[11] *B. subtilis* has unique properties such as spore formation, versatility of growth nutrients utilization, high level of enzymes production, fast growth rate, and growth in aerobic and anaerobic conditions.^[3]

Synthesis of antimicrobial agents

Bacillus bacteria play a significant role in the gut because of their high metabolic activity. The activity of Bacillus is mainly determined by their ability to produce antibiotics. B. subtilis is the most productive species which devotes 4%-5% of genome to antibiotic synthesis and produce 66 antibiotics. Bacillus antibiotics have different structure and spectrum of antimicrobial activity.^[5] The potential of *B. subtilis* to produce antibiotics has been recognized for 50 years. Antimicrobial agents synthesized and secreted by B. subtilis are listed in Table I.^[12-14] B. subtilis synthesized antimicrobial substances have antimicrobial effects against broad spectrum of pathogens. These substances are natural part of human antimicrobial defense system hence the possibility of developing pathogen resistance or unwanted side effects is less. Hence, probiotic B. subtilis is ideal therapeutic option because of their broad spectrum of activity and specific and rapid killing activity against various pathogens.^[14] B. subtilis probiotic properties are strain-specific.^[3] Pectinolytic enzymes have been isolated from B. subtilis. Bacilli produce amino acids, including essential amino acids and vitamins.^[5]

Immunostimulatory effect

B. subtilis enhances the protection against pathogens by stimulating nonspecific and specific immunity. A number of studies in humans and animal models have provided strong evidence that oral administration of Bacillus spores stimulates the immune system. Spores of B. subtilis trigger specific humoral and cell-mediated immune responses. The interaction between B. subtilis spores and macrophages plays an important role in development of both innate and adaptive immune responses of the host. Numerous studies have demonstrated that B. subtilis leads to macrophage activation. B. subtilis B10, B. subtilis BS02, and B. subtilis (natto) B4 spores may possess immunomodulatory activities by the induction of pro-inflammatory cytokines and exerts probiotic activities through activated macrophage functions. In addition, B. subtilis showed no apparent cytotoxicity against RAW 264.7 cells and thought to be safe.^[15-17] B. subtilis MBTU PBBM1 spores administration leads to augmentation of antibody response (antibodies IgG and IgA) and also proliferation of the T lymphocytes which indicates B. subtilis MBTU PBBM1 spores have the potential to improve both the humoral and cellular immunity in mice.^[18] Commensal bacteria play an important role in the development of the gut-associated

lymphoid tissue (GALT) and important for both innate and adaptive immunity. B. subtilis promotes active lymphocyte proliferation within GI tract. B. subtilis administration in appendix of germfree rabbits has been shown to promote GALT development. This evidence showed that Bacillus species are important for development of robust gut-associated lymphoid system (GALT) and promote a potent immune response.^[19] The effect of lymphocyte activation by *B. subtilis* spores was both quantitatively and qualitatively similar to mitogens phytohaemagglutinin and Concanavalin A. B. subtilis spores stimulated cytokine production in vitro and after oral administration in mice.^[5] Oral treatment with *B. subtilis* spores increased expression of activation markers on lymphocytes in dose-dependent manner in healthy volunteers.^[20] B. subtilis spores-induced systemic antibody response to tetanus toxoid fragment C and ovalbumin in mice. These data suggest that B. subtilis spores are an efficient mucosal and systemic adjuvant for enhancing humoral immune responses.^[21]

Maintenance of intestinal homeostasis and prevention of intestinal inflammation

B. subtilis derived quorum-sensing pentapeptide, competence and sporulation factor (CSF) activate key survival pathways including p38 MAP kinase and protein kinase B (Akt) in intestinal epithelial cells of the host. CSF also induces heat shock proteins, which protect intestinal epithelial cells against injury and loss of barrier function and hence provide the ability to maintain intestinal homeostasis.^[22] B. subtilis quorum sensing molecule CSF reduced epithelial injury caused by intestinal inflammation and improved the survival rate of mice with lethal colitis. This indicates that B. subtilis are potentially useful for treating intestinal inflammation. B. subtilis is beneficial for maintaining intestinal homeostasis and host health and can be utilized to treat antibiotics-induced colitis, inflammatory bowel disease (IBD) (including Crohn's disease and ulcerative colitis) and necrotizing enterocolitis.^[23] Bacillus species (B. subtilis, Bacillus firmus, Bacillus megaterium, and B. pumilus) have been shown to convert genotoxic compounds to unreactive products in vitro.^[24] Orally administered B. subtilis spores were effective in decreasing infection and enteropathy in suckling mice infected with Citrobacter rodentium (a model for the traveler's diarrhea pathogen enterotoxigenic Escherichia coli) which is known to cause epithelial lesions, crypt hyperplasia, and mortality.^[7,25] Intestinal mucosal barrier dysfunction associated with IBD. Mucosal biopsies taken from IBD patients showed loss of key epithelial tight junction (TJ) proteins such as claudin-I, occludin, junctional adhesion molecule (JAM)-A, and zona occludens (ZO)-1. Effects of B. subtilis on epithelial TJs and intrinsic regulatory mechanisms of the intestine were studied in dextran sulfate sodium-induced colitis in Balb/c mice. B. subtilis significantly reduced disease activity index scores and graded histologic damage. B. subtilis improved

Category of antimicrobial agents	Antimicrobial agents	Mechanism of action	Spectrum of antimicrobial activity
Ribosomal synthesized peptides Bacteriocins: Type A lantibiotics	Subtilin, ericin A, ericin S	Voltage-dependent pores into the cytoplasmic membrane	Gram-positive bacteria, antiviral, and antimycoplasma activities
Ribosomal synthesized peptides: Type B lantibiotics	Mersacidin	Inhibition of cell wall synthesis	Gram-positive bacteria, including methicillin-resistant strains of <i>S. aureus</i> and vancomycin resistant strains of <i>Enterococc</i>
Unusual lantibiotics	Subtilosin A	Antimicrobial activity by interacting with membrane-associated receptors	Gram-positive bacteria, strong bactericidal activity against <i>L. monocytogenes</i>
	Sublancin 168	-	Gram-positive bacteria, pathogens such as <i>B. cereus</i> , <i>S. pyogenes</i> and <i>S. aureus</i>
	Bacillocin 22	-	-
Nonribosomal synthesized peptides	Surfactin	Powerful biosurfactant - it exerts a detergent-like action on biological membranes	Antiviral and antimycoplasma activities, inhibit biofilm formation of human pathogen <i>S. enterica</i>
	Bacilysin	Inhibits glucosamine synthase involved in synthesis of nucleotides, amino acids and coenzymes and resulting in lysis of microbial cells	S. aureus and C. albicans
	Iturine lipopeptides: Mycosubtilin, iturines,		Strong antifungal and hemolytic and limited antibacterial activities
	bacillomycins		Antimiarahial and funciaidal action
	Fengycin (plipastatin)		Antimicrobial and fungicidal action
	Rhizocticins Mycobacillin (B3)		Antifungal activity Antibacterial and antifungal activities
	Corynebactin (bacillibactin),	-	
	3,3'-neotrehalosadiamine (168),		
	difficidin, TL-119 (A-3302-B)		
Miscellaneous	Polyketides: Difficidin,	-	Antibacterial activity against both aerobic and anaerobic
antibiotic compounds	oxydifficidin, bacillaene		organisms
	Amicoumacin	-	Antibacterial activities, S. aureus and H. pylori
	Bacilysocin	-	-
	BSAP-254	-	Antagonistic effect against the food borne pathogens
	Entianin	-	Strong antibacterial activity against <i>S. aureus, E. faecalis,</i> and other Gram-positive pathogens
	Subpeptin JM4-A, subpeptin	-	Active against broad spectrum of bacteria, including
	JM4-B Antifungal protein B29I	-	Salmonella, B. cereus, S. aureus Inhibitory activity on mycelial growth in F. oxysporum, Rhizoctonia solani and other fungi
	Bacteriocin-like substances		Gram-positive and Gram-negative bacteria
	AMP IC-1	-	Antagonistic to <i>B. cereus</i>
	AMPNT-6	-	Active against marine food borne pathogen
			Useful for seed disinfection

Table 1: Antimicrobial agents synthesized and secreted by Bacillus subtilis

-: Not yet identified, S. aureus: Staphylococcus aureus, L. monocytogenes: Listeria monocytogenes, S. pyogenes: Streptococcus pyogenes, S. enterica: Salmonella enterica, C. albicans: Candida albicans, H. pylori: Helicobacter pylori, E. faecalis: Enterococcus faecalis, B. cereus: Bacillus cereus, F. oxysporum: Fusarium oxysporum, R. solani: Rhizoctonia solani

barrier function by upregulating expression of epithelial TJs proteins (claudin-I, occludin, JAM-A, and ZO-I) and reduced intestinal epithelial damage by downregulating cytokine expression (interleukin-6 [IL-6], IL-17, IL-23, and tumor necrosis factor- α).^[26]

Maintenance of intestinal normal flora

B. subtilis positive effect on the maintenance of the normal intestinal flora has been demonstrated in many studies. *B. subtilis* 3 strain showed efficacy against pathogenic cultures of *E. coli* and *Campylobacter* species during treatment of

experimental infections in mice and maintained normal microflora in the animals during receipt of antibiotic therapy. In vitro studies of B. subtilis, 3 showed a wide spectrum of antagonistic activity toward the tested pathogens and did not inhibit normal microflora.^[27] B. subtilis MA139 significantly increased the number of Lactobacillus and reduced the content of E. coli in the intestines and feces in pigs.^[28] B. subtilis KN-42 significantly increased lactobacilli counts and reduced E. coli counts and improved the growth performance and GI health of piglets.^[29] B. subtilis KDI improved intestinal flora by significantly increasing lactobacilli counts and reducing E. coli counts and improve the growth performance in broilers.^[30] B. subtilis var. natto in mice influenced the fecal microflora, specifically Bacteroides and Lactobacillus species. Mice fed with an egg white diet showed decrease in numbers of Lactobacillus spp., while B. subtilis var. natto spores supplemented diet stabilized it. Using a casein diet, the numbers of Bacteroidaceae increased. This result indicated that B. subtilis var. natto could be beneficial in maintaining the natural microflora.^[24] Therefore, an increase in Lactobacillus counts and decrease in E. coli counts may result in a lower diarrhea incidences.^[29] Salmonella is a major foodborne pathogen which can cause severe illness in humans such as enteric fever, bacteremia, focal infection, and enterocolitis. B. subtilis NCII exhibits strong inhibition activity against Salmonella enteritidis infection to intestinal epithelial cells.^[31] B. subtilis CUI effects on intestinal mucosal immune system, and microbial balance were evaluated in antibiotic-induced dysbiosis mouse model. B. subtilis CUI spores $(3 \times 10^9 \text{ spores/day/mouse})$ administered before and during the antibiotic treatments. Treatment with the B. subtilis CUI strain decreases the antibiotic-induced intestinal inflammation. B. subtilis CUI shown to normalize the B220+MHCII+B-cells in mesenteric lymphoid node and F4/80+ macrophages in Peyer's patches in the antibiotic group. B. subtilis CUI treatment reduced antibiotic-induced alterations in the gut microbiota. This result suggests that B. subtilis CUI may contribute to the reduction of antibiotic-induced inflammation through normalization of mucosal immune responses and intestinal microbiota.^[32]Taking into account beneficial properties of B. subtilis, this bacterium is a potential probiotic candidate to be considered for various clinical conditions.

CLINICAL TRIALS OF BACILLUS SUBTILIS

B. subtilis therapy was highly effective in treatment of various infectious pathologies in patients.^[33] Clinical efficacy of *B. subtilis* was summarized as an antidiarrheal agent, used in different counties. *B. subtilis* is one of the most important microorganisms for the treatment and prophylaxis of intestinal disorders in humans.^[34] *B. subtilis* was more effective in treatment of acute diarrhea than lactobacilli.^[5]

Regularity of bowel movements

Labellarte *et al.*, carried out randomly assigned, double-blind placebo-controlled trial of *B. subtilis* (approximately 1.9×10^9 CFU/capsule) in 40 healthy male and female adults for an average of 20 days. The study showed that daily consumption of *B. subtilis* was effective in promoting regularity of bowel movements and well tolerated.^[35]

Survival in the gastrointestinal tract

Hanifi *et al.*, carried out randomized, double-blind, placebo-controlled trial of *B. subtilis* R0179 in 81 healthy adults (18–50 years old). Subjects received *B. subtilis* R0179 at dose of 0.1, 1 or 10 × 10⁹ CFU/capsule/day or placebo for 4 weeks. Fecal viable counts of *B. subtilis* R0179 showed a dose-dependent GI survival response and fecal viable counts were 0.1 × 10⁹ (4.6 ± 0.1 log₁₀ CFU/g), 1 × 10⁹ (5.6 ± 0.1 log₁₀ CFU/g) and 10 × 10⁹ (6.4 ± 0.1 log₁₀ CFU/g) respectively (P < 0.0001). *B. subtilis* R0179 survives passage through the human GI tract and is safe and well tolerated in healthy adults at intake from 0.1 to 10 × 10⁹ CFU/day.^[36]

Diarrhea and antibiotic-associated diarrhea

Clinical efficacy of Bacillus bacteria in the treatment of GI infections has been reported. Mazza (1994) summarized results of numerous studies and concluded that B. subtilis are one of the most important microorganisms for the treatment and prophylaxis of intestinal disorders in humans.^[34] In clinical study, B. subtilis and B. licheniformis (2×10^9) microbial cells; Biosporin) were administered to the patients with acute enteric infections. Results showed the pronounced curative effect of Bacillus probiotics manifested by the rapid normalization of stool, abdominal pain relief, and decrease in intestinal dysbiosis. Bacillus probiotics found to be safe and well tolerated.^[37] B. subtilis and B. licheniformis (Biosporin) has been also evaluated for effect on intestine microflora in acute digestive disorders and dysbacterioses in 53 newborn children with perinatal pathology. Results showed high therapeutic and prophylactic efficiency for dysbacterioses and diarrheas in the newborn children without side effects.^[38] One of the most common side effects of antibiotic therapy is antibiotic-associated diarrhea (AAD). The frequency of AAD depends on the type of antibiotic used and varies from 25% to as high as 44%. The route of antibiotic administration (oral or parenteral) does not affect the rate of AAD, and no difference has been found in the frequency of AAD with respect to age and gender. The severity of AAD may vary from uncomplicated diarrhea to Clostridium difficile-associated pseudomembranous colitis. The main mechanism for the development of AAD is significant changes in the composition and quantity of the gut microbiota during the treatment with antibiotics. AAD may be caused by different enteric pathogens such as Salmonella spp., Staphylococcus aureus, Candida albicans, Clostridium perfringens, and Klebsiella spp. Bacillus bacteria have attracted the growing attention of researchers as effective probiotics for the treatment and prevention of enteric infections. Research study showed high efficacy of the Bacillus probiotic B. subtilis 3 and B. licheniformis 31 (predominant amount of B. subtilis 3 in 50:1 parts; Biosporin) in the treatment of acute intestinal infections.^[39] In clinical trial, B. subtilis spores (4×10^9) administered to 11 children aged 3-24 months for 5 days along with antibiotic and alone antibiotic was given to 8 subjects. Results showed that number of stools increased in alone antibiotic group while in B. subtilis along with antibiotic group no such changes were observed. Bacteriotherapy along with antibiotic also increased saccharolytic flora, aerobic and anaerobic flora.^[40] Horosheva et al. carried out a randomized, double-blind, placebo-controlled clinical trial on outpatients aged \geq 45 years who were prescribed \geq 1 oral or intravenous antibiotics for at least 5 days. One group of patients (n = 90) received probiotic B. subtilis 3 (2×10^9 CFU/vial), 2 times a day from beginning I day before initiation of antibiotic therapy and ending 7 days after discontinuation of antibiotic. Results showed that AAD developed in 25.6% (23/90) patients in placebo group while in significantly lower AAD rate 7.8% (7/90) patients reported in B. subtilis 3 group (P < 0.001). B. subtilis 3 significantly reduced the incidence of nausea, vomiting, bloating, and abdominal pain.^[39] There have been 23 clinical trials involving over 1800 patients for probiotic preparation containing a combination of B. subtilis R0179 and other probiotic. It has been used in the improvement of symptoms associated with chronic diarrhea and irritable bowel syndrome, as a coadjuvant therapy with sulfasalazine and mesalazine to improve remission times in mild to moderate ulcerative colitis and to improve compliance with conventional triple therapy for Helicobacter pylori eradication.[41]

Stimulation of immune responses

Lefevre et al. (2015) carried out randomized, double-blind, placebo-controlled, parallel-arms trial on 100 elderly subjects



SAFETY OF BACILLUS SUBTILIS

According to European Scientific Committee on Animal Nutrition, *B. subtilis* was tested and showed no evidence of toxicity. Acute and chronic toxicity studies in animals also indicated safety of these strains. *B. subtilis* is generally recognized as safe by the Food and Drug Administration (FDA), meaning it is not harmful to humans.^[3] *B. subtilis* species is considered safe by the European Food Safety Authority (EFSA) Qualified Presumption of Safety. Thus, *B. subtilis* strain may be considered as nonpathogenic and safe for human consumption.^[43] *B. subtilis* could be considered as a perfect multifunctional probiotic bacterium for humans.^[3]



Figure 2: Increase in secretory IgA level in saliva

Figure 3: Increase in secretory IgA level in stools

Day 10

P+0.0038

Bacillus subtilis CU1

After 18 days break

P=0.0032 (N=44)

SUMMARY

The world market for probiotics supplements has been growing over the last two decades based on their important clinical merits. Lactobacillus and Bifidobacterium are the most used genera, mainly for their ability to exclude pathogens. However, they do not have multifunctional probiotic capacities as B. subtilis. Bacillus bacteria are increasingly attracting attention of researchers as promising probiotics due to their strong antimicrobial, antidiarrheal and immunostimulatory effects, ability to stimulate growth of natural flora and prevent intestinal inflammation; besides having an excellent stability profile in otherwise unfavorable conditions. Moreover, it has established efficacy and safety in numerous randomized, double-blind clinical trials, as validated and approved by authorities like FDA and EFSA. In this sense, B. subtilis has the potential to emerge as the "perfect multifunctional probiotic bacteria" for various clinical conditions in humans.

Financial support and sponsorship Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- 1. Reid G, Jass J, Sebulsky MT, McCormick JK. Potential uses of probiotics in clinical practice. Clin Microbiol Rev 2003;16:658-72.
- FAO/WHO. Probiotic in Foods: Health and Nutritional Properties and Guidelines for Evaluation. In FAO Food and Nutrition. Rome, Italy: FAO/ WHO; 2005. p. 85.
- Olmos J, Paniagua-Michel J. *Bacillus subtilis* a potential probiotic bacterium to formulate functional feeds for aquaculture. J Microb Biochem Technol 2014;6:7.
- Vasquez AP. Bacillus species are superior probiotic feed-additives for poultry. J Bacteriol Mycol Open Access 2016;2:00023.
- Sorokulova I. Modern status and perspectives of *Bacillus* bacteria as probiotics. J Prob Health 2013;1:4.
- Urdaci MC, Bressollier P, Pinchuk I. *Bacillus clausii* probiotic strains: Antimicrobial and immunomodulatory activities. J Clin Gastroenterol 2004;38 6 Suppl: S86-90.
- 7. Cutting SM, Van PH, Dong TC. Bacillus probiotics. Nutra foods 2009;8:7-14.
- Tam NK, Uyen NQ, Hong HA, Duc le H, Hoa TT, Serra CR, et al. The intestinal life cycle of *Bacillus subtilis* and close relatives. J Bacteriol 2006;188:2692-700.
- Casula G, Cutting SM. *Bacillus* probiotics: Spore germination in the gastrointestinal tract. Appl Environ Microbiol 2002;68:2344-52.
- Ozawa K, Yokota H, Kimura M, Mitsuoka T. Effects of administration of Bacillus subtilis strain BN on intestinal flora of weanling piglets. Nihon Juigaku Zasshi 1981;43:771-5.
- Leser TD, Knarreborg A, Worm J. Germination and outgrowth of *Bacillus* subtilis and *Bacillus licheniformis* spores in the gastrointestinal tract of pigs. J Appl Microbiol 2008;104:1025-33.
- Stein T. Bacillus subtilis antibiotics: Structures, syntheses and specific functions. Mol Microbiol 2005;56:845-57.
- Baruzzi F, Quintieri L, Morea M, Caputo L. Antimicrobial compounds produced by *Bacillus* spp. and applications in food. In: Vilas AM, editor. Science against Microbial Pathogens: Communicating Current Research and Technological Advances. Badajoz, Spain: Formatex; 2011. p. 1102-11.

- Sumi CD, Yang BW, Yeo IC, Hahm YT. Antimicrobial peptides of the genus Bacillus: A new era for antibiotics. Can J Microbiol 2015;61:93-103.
- Huang Q, Xu X, Mao YL, Huang Y, Rajput IR, Li WF. Effects of *Bacillus* subtilis B10 spores on viability and biological functions of murine macrophages. Anim Sci J 2013;84:247-52.
- Huang Q, Li YL, Xu X, Huang Y, Cui ZW, Yu DY, et al. Modulatory effects of Bacillus subtilis BS02 on viability and immune responses of RAW 264.7 murine macrophages. J Anim Vet Adv 2012;11:1934-8.
- Xu X, Huang Q, Mao Y, Cui Z, Li Y, Huang Y, *et al.* Immunomodulatory effects of *Bacillus subtilis (natto)* B4 spores on murine macrophages. Microbiol Immunol 2012;56:817-24.
- Sebastian AP, Keerthi TR. Immunomodulatory effect of probiotic strain Bacillus subtilis MBTU PBBMI spores in Balb/C Mice. Int J Eng Tech Res 2014;2:258-60.
- Huang JM, La Ragione RM, Nunez A, Cutting SM. Immunostimulatory activity of *Bacillus* spores. FEMS Immunol Med Microbiol 2008;53:195-203.
- Caruso A, Flamminio G, Folghera S, Peroni L, Foresti I, Balsari A, *et al.* Expression of activation markers on peripheral-blood lymphocytes following oral administration of *Bacillus subtilis* spores. Int J Immunopharmacol 1993;15:87-92.
- Barnes AG, Cerovic V, Hobson PS, Klavinskis LS. *Bacillus subtilis* spores: A novel microparticle adjuvant which can instruct a balanced Th1 and Th2 immune response to specific antigen. Eur J Immunol 2007;37:1538-47.
- Fujiya M, Musch MW, Nakagawa Y, Hu S, Alverdy J, Kohgo Y, *et al.* The Bacillus subtilis quorum-sensing molecule CSF contributes to intestinal homeostasis via OCTN2, a host cell membrane transporter. Cell Host Microbe 2007;1:299-308.
- Okamoto K, Fujiya M, Nata T, Ueno N, Inaba Y, Ishikawa C, *et al.* Competence and sporulation factor derived from *Bacillus subtilis* improves epithelial cell injury in intestinal inflammation via immunomodulation and cytoprotection. Int J Colorectal Dis 2012;27:1039-46.
- Hong HA, Duc Le H, Cutting SM. The use of bacterial spore formers as probiotics. FEMS Microbiol Rev 2005;29:813-35.
- D'Arienzo R, Maurano F, Mazzarella G, Luongo D, Stefanile R, Ricca E, et al. Bacillus subtilis spores reduce susceptibility to *Citrobacter* rodentium-mediated enteropathy in a mouse model. Res Microbiol 2006;157:891-7.
- Gong Y, Li H, Li Y. Effects of *Bacillus subtilis* on epithelial tight junctions of mice with inflammatory bowel disease. J Interferon Cytokine Res 2016;36:75-85.
- Sorokulova I. Preclinical testing in the development of probiotics: A regulatory perspective with *Bacillus strains* as an example. Clin Infect Dis 2008;46 Suppl 2:S92-5.
- Guo X, Li D, Lu W, Piao X, Chen X. Screening of *Bacillus* strains as potential probiotics and subsequent confirmation of the *in vivo* effectiveness of *Bacillus* subtilis MA139 in pigs. Antonie Van Leeuwenhoek 2006;90:139-46.
- Hu Y, Dun Y, Li S, Zhao S, Peng N, Liang Y. Effects of *Bacillus subtilis* KN-42 on growth performance, diarrhea and faecal bacterial flora of weaned piglets. Asian Australas J Anim Sci 2014;27:1131-40.
- Wu BQ, Zhang T, Guo LQ, Lin JF. Effects of *Bacillus subtilis* KD1 on broiler intestinal flora. Poult Sci 2011;90:2493-9.
- Thirabunyanon M, Thongwittaya N. Protection activity of a novel probiotic strain of *Bacillus subtilis* against *Salmonella enteritidis* infection. Res Vet Sci 2012;93:74-81.
- Racedo S, Jacquot C, Pinchuk I, Urdaci M. F. 84. Probiotic *Bacillus subtilis* CU1 Normalize the Mucosal Immune Responses and Microbiota Balance in a Murine Model of Dysbiosis. 15th International Congress of Mucosal Immunology (ICMI 2011) Abstract Supplement; 5-9 July, 2011. p. 151.
- Pham Ngoc T, Vu Thi C, Nguyen Thi H. Bacillus subtilis in therapy and prevention of disease. Rev Immunol Ther Antimicrob 1968;32:53-65.
- Mazza P. The use of *Bacillus subtilis* as an antidiarrhoeal microorganism. Boll Chim Farm 1994;133:3-18.
- Labellarte G, Cooper S, Maher M. Tolerance and efficacy of a probiotic supplement delivered in capsule form. FASEB J 2015;29 Suppl 924:33.
- Hanifi A, Culpepper T, Mai V, Anand A, Ford AL, Ukhanova M, et al. Evaluation of *Bacillus subtilis* R0179 on gastrointestinal viability and general wellness: A randomised, double-blind, placebo-controlled trial in healthy adults. Benef Microbes 2015;6:19-27.

- Gracheva NM, Gavrilov AF, Solov'eva AI, Smirnov VV, Sorokulova IB, Reznik SR, *et al.* The efficacy of the new bacterial preparation biosporin in treating acute intestinal infections. Zh Mikrobiol Epidemiol Immunobiol 1996;(1):75-7.
- Slabospitskaia AT, Vinogradov VP, Krymovskaia SS, Reznik SR, Smirnov VV. A new preparation of biosporin and its effect on the intestinal microflora in dysbacterioses in newborn infants. Mikrobiol Z 1995;57:71-6.
- Horosheva TV, Vodyanoy V, Sorokulova I. Efficacy of *Bacillus* probiotics in prevention of antibiotic-associated diarrhoea: A randomized, double-blind, placebo-controlled clinical trial. JMM Case Rep 2014;1:1-6.
- 40. Benoni G, Marcer V, Cuzzolin L, Raimo F. Antibiotic administration and oral

bacterial therapy in infants. Chemioterapia 1984;3:291-4.

- Tompkins TA, Xu X, Ahmarani J. A comprehensive review of post-market clinical studies performed in adults with an Asian probiotic formulation. Benef Microbes 2010;1:93-106.
- Lefevre M, Racedo SM, Ripert G, Housez B, Cazaubiel M, Maudet C, *et al.* Probiotic strain *Bacillus subtilis* CU1 stimulates immune system of elderly during common infectious disease period: A randomized, double-blind placebo-controlled study. Immun Ageing 2015;12:24.
- Sorokulova IB, Pinchuk IV, Denayrolles M, Osipova IG, Huang JM, Cutting SM, *et al.* The safety of two *Bacillus* probiotic strains for human use. Dig Dis Sci 2008;53:954-63.

